Interactions of selenium hyperaccumulators and nonaccumulators during cocultivation on seleniferous or nonseleniferous soil – the importance of having good neighbors

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Summary

• This study investigated how selenium (Se) affects relationships between Se hyperaccumulator and nonaccumulator species, particularly how plants influence their neighbors’ Se accumulation and growth.

• Hyperaccumulators Astragalus bisulcatus and Stanleya pinnata and nonaccumulators Astragalus drummondii and Stanleya elata were cocultivated on seleniferous or nonseleniferous soil, or on gravel supplied with different selenate concentrations. The plants were analyzed for growth, Se accumulation and Se speciation. Also, root exudates were analyzed for Se concentration.

• The hyperaccumulators showed 2.5-fold better growth on seleniferous than on nonseleniferous soil, and up to fourfold better growth with increasing Se supply; the nonaccumulators showed the opposite results. Both hyperaccumulators and nonaccumulators could affect growth (up to threefold) and Se accumulation (up to sixfold) of neighboring plants. Nonaccumulators S. elata and A. drummondii accumulated predominantly (88–95%) organic C-Se-C; the remainder was selenate. S. elata accumulated relatively more C-Se-C and less selenate when growing adjacent to S. pinnata. Both hyperaccumulators released selenocompounds from their roots. A. bisulcatus exudate contained predominantly C-Se-C compounds; no speciation data could be obtained for S. pinnata.

• Thus, plants can affect Se accumulation in neighbors, and soil Se affects competition and facilitation between plants. This helps to explain why hyperaccumulators are found predominantly on seleniferous soils.

Introduction

Selenium (Se) is an essential element for humans and animals, but it is toxic at higher concentrations (Terry et al., 2000). There is a narrow margin between Se deficiency and toxicity in animals (Stadtman, 1990). As an essential element, Se is required for the production of selenoproteins, some of which function in scavenging free radicals (Zhang & Gladyshev, 2009). Selenium deficiency may promote cancer and cause other diseases such as white muscle disease, which may be fatal (Cosgrove, 2001). Selenium toxicity is thought to be the result of the similarity of Se to sulfur (S); substitution of S by Se in proteins disrupts protein structure and function (Stadtman, 1990). For plants, Se is also toxic at high concentrations (Anderson, 1993). The essentiality of Se for higher plants is still unproven, but Se is considered a beneficial nutrient for many plant species (Pilon-Smits et al., 2009), perhaps because of better oxidative stress resistance (Cartes et al., 2005; Hartikainen, 2005). Plants readily take up and assimilate Se, a capacity that may be used to alleviate both Se deficiency and toxicity in animals and humans. Plants can be used to clean up excess Se from polluted areas (phytoremediation), and Se-enriched plant material may be considered fortified food (biofortification) (Terry et al., 2000).

Plants mainly take up Se from soil in the form of selenate (SeO₄²⁻), which is taken up inadvertently via sulfate transporters, and metabolized via the S assimilation pathway (for a review see Sors et al., 2005). In this pathway, selenite is reduced to selenide (SeO₂⁻), which can undergo further reduction to selenide (Se⁻²). This may be incorporated into the organic forms, selenocysteine (SeCys), selenocystathionine (SeCysth) and selenomethionine (SeMet). Plant species differ in their capacity to accumulate Se. While most plant species accumulate Se to concentrations below 100 mg Se kg⁻¹ DW, even when growing on Se-rich (seleniferous) soils, some plant species native to seleniferous soils can accumulate Se to concentrations as high as 10 000 mg Se kg⁻¹ DW (Beath et al., 1934, 1939; Galeas et al.,...
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concentrations in the soil were high enough to explain the medium supplied with different concentrations of Se, the Se
et al. Arabidopsis thaliana lower germination and growth of the Se-sensitive model plant soil collected next to hyperaccumulators yielded significantly 10% lower around hyperaccumulators than around comparable neighbors may have an allelopathic effect if they are Se-sensitive. When taken to the laboratory and used in controlled herbivory studies with grasshoppers collected from the same field site, the high-Se S. ericoides and A. ludoviciana plants collected next to hyperaccumulators were eaten less than their low-Se counterparts collected next to nonaccumulators (El Mehdawi et al., 2011b).

Several questions remain regarding the effects of hyperaccumulated Se on plant–plant interactions. First, is the higher Se concentration in soil around hyperaccumulators a result of litter deposition or root exudation, or both? In an earlier study it was found that high-Se litter decomposed readily in a seleniferous habitat, harbored more microbial and micro-arthropod decomposers than low-Se litter, and led to enrichment of the underlying soil with Se (Quinn et al., 2010). Release of Se by hyperaccumulator roots via exudation and turnover has never been tested, but may also be substantial, since hyperaccumulator roots can contain Se concentrations around 0.3% of DW (Galeas et al., 2007). Secondly, are the higher Se concentrations in neighboring plants solely a result of higher total soil Se concentrations, or also the result of different chemical Se speciation in soil around hyperaccumulators (which may affect Se bioavailability), and/or of the presence of Se chelators? Do hyperaccumulators affect the speciation of Se in their local environment, including neighboring plants? Thirdly, how is the competition between hyperaccumulators and nonhyperaccumulators affected by the Se concentration of the soil? In this study we aim to address these questions.

Materials and Methods

Soil collection and characterization

Soil was collected in June 2010 from two sites on the West side of Fort Collins, CO, USA: Pine Ridge Natural Area (40°32.70N, 105°07.87W, elevation 1510 m), and Cloudy Pass (40°37.33N, 105°12.38W, elevation 1570 m). Pine Ridge Natural Area is a seleniferous area with soil composed of Se-rich Cretaceous shale. This semi-arid shrubland harbors at least two species of Se-hyperaccumulating plants: Astragalus bisulcatus (Hook.) A. Gray and Stanleya pinnata (Pursh) Britton (Galeas et al., 2007). Cloudy Pass is a nonseleniferous area 10 miles northwest of Pine Ridge Natural Area and similar in altitude, climate and vegetation except that no Se hyperaccumulators are present. Cloudy Pass does contain the nonhyperaccumulator species Astragalus drummondii Douglas ex Hook. Soil samples were collected at both sites from 0 to 5 cm depth to determine soil properties and elemental concentrations. Soil pH and electroconductivity (EC) were determined as described using a saturated soil paste (Soil Survey Laboratory Methods Manual, 2004). Soil

Interestingly, in some cases, hyperaccumulators can also have a positive effect on their plant neighbors (facilitation), if these neighbors are Se-resistant. In field studies, Symphyotrichum ericoides and Artemisia ludoviciana were twofold bigger when growing next to hyperaccumulators than when they were growing next to nonaccumulators (El Mehdawi et al., 2011b). This benefit appeared to be, at least in part, the result of enhanced protection from herbivory: S. ericoides and A. ludoviciana harbored fewer herbivores in the field and exhibited less herbivory. Moreover, since hyperaccumulators are found predominantly on seleniferous soils, they appear to have a physiological or ecological need for Se (Beath et al., 1934). There is ample evidence that Se serves ecological functions for hyperaccumulators. Selenium has been shown to protect plants from a wide variety of invertebrate and vertebrate herbivores, as a result of a combination of deterrence and toxicity (Hurd-Karrer & Poos, 1936; Vickerman et al., 2002; Hanson et al., 2003, 2004; Freeman et al., 2006b, 2007, 2009; Galeas et al., 2008; Quinn et al., 2008, 2010). In addition to herbivores, Se can protect plants from Se-sensitive fungal pathogens (Hanson et al., 2003). Thus, hyperaccumulators may have an ecological dependency on Se for protection from various biotic stresses. In addition to elemental defense, Se may be used by hyperaccumulators for elemental allelopathy. Soil adjacent to hyperaccumulators was 7- to 13-fold enriched in Se compared with plants from the same species growing > 4 m away from hyperaccumulators (El Mehdawi et al., 2011a,b). Accordingly, neighboring vegetation of hyperaccumulators contained two- to 20-fold elevated Se concentrations compared with plants from the same species growing > 4 m away from hyperaccumulators (El Mehdawi et al., 2011a,b). The higher Se concentrations in neighbors of hyperaccumulators may have an allelopathic effect if they are Se-sensitive. Indeed, the percentage vegetative ground cover was on average 10% lower around hyperaccumulators than around comparable nonaccumulator species (El Mehdawi et al., 2011a). Moreover, soil collected next to hyperaccumulators yielded significantly lower germination and growth of the Se-sensitive model plant Arabidopsis thaliana, and higher Se accumulation, than soil collected around nonhyperaccumulator species (El Mehdawi et al., 2011a). Based on controlled experiments using agar medium supplied with different concentrations of Se, the Se concentrations in the soil were high enough to explain the observed inhibitive effect on A. thaliana germination (El Mehdawi et al., 2011a).

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texture was determined as described by Gee & Bauder (1986) using a hydrometer method for sand, silt and clay. Soil organic matter (SOM) was determined using a modification of the Walkley Black method, by means of a Spectronic 20 (Milton Roy Co., Ivyland, PA, USA) at 610 nm (Soltanpour & Workman, 1981). Soil calcium carbonate (CaCO₃) was quantified using gravimetric determination from CO₂ evolution (Soil Survey Laboratory Methods Manual, 2004). Soil elemental analysis was performed as described in the following.

Plant material

Seeds from *A. bisulcatus* and *A. drummondii* were obtained from Western Native Seed, Coaldale, CO, USA. *Stanleya pinnata* seeds were collected from seleniferous soil at Pine Ridge Natural Area in Fort Collins, CO. *Stanleya elata* M.E. Jones seeds (accession #113) were collected from nonseleniferous soil in Nevada at N 37°26.699′ W 117°21.896′, at an elevation of 1515 m.

Cocultivation experiment on seleniferous and nonseleniferous soils

The soil collected from Pine Ridge Natural Area (seleniferous) and from Cloudy Pass (nonseleniferous) was sieved (1 mm mesh) to remove large stones and organic material, and mixed 3 : 1 with Turface® (Buffalo Grove, IL, USA) to make the aeration adequate and to enhance drainage. A thin layer of course gravel and sand was placed in the bottom of 10 × 10 cm pots, and the soil–Turface mixture placed on top. Each pot was placed on an individual tray to catch leachate and keep it available for the plants.

*Stanleya pinnata* and *S. elata* seeds were surface-sterilized by rinsing for 20 min in 20% bleach, followed by five 10 min rinses in sterile water. The *A. bisulcatus* and *A. drummondii* seeds were first scarified with sand paper and then surface-sterilized. The seeds were germinated on sterilized, wet filter paper under continuous light at 23°C in a plant growth cabinet. The emerging seedlings were carefully transferred to the pots. Two plants were placed in each pot. For each soil type the following seven species combinations were created, using six replicates per treatment: two plants of the same species, from *A. bisulcatus*, *S. pinnata*, *A. drummondii* or *S. elata*; two plants of different species, either one hyperaccumulator and one nonhyperaccumulator (*A. bisulcatus* and *A. drummondii* or *S. pinnata* and *S. elata*); or two hyperaccumulators (*A. bisulcatus* and *S. pinnata*). The plants were watered twice a wk with water and once a wk with 0.5-strength Hoagland solution (Hoagland & Arnon, 1938). After 2 months, when the plants became bigger, they were transferred to 14-cm-diameter round pots. Again, a thin layer of gravel and sand was placed on the bottom, and the area around the transplanted soil was filled up with a similar mixture of soil (from the same source as originally) and Turface. The plants were harvested after 6 months, gently washed, and transferred to 50 ml of distilled water. After 2 d the plants were transferred to another container with 50 ml water. After another 3 d, this second volume of water and root-released compounds (which will hereafter be referred to as exudate) was collected and analyzed for Se concentration as described in the following. Furthermore, some of the exudate fractions were frozen for Se speciation as described later. In addition, the exudate fractions were used to extract some Pine Ridge Natural Area soil. To 2 g of soil was added 6 ml of exudate, and after mixing by rotation for 1 h at room temperature, allowed to settle overnight at 4°C. The liquid fraction was then removed and used for elemental analysis and X-ray microprobe analyses as described later.

Selenium distribution and speciation

Selenium speciation was compared in leaf material of *S. elata* grown next to *S. elata* and *S. elata* grown next to *S. pinnata*, as well as in leaves of *A. drummondii* growing next to *A. drummondii* and *A. drummondii* growing next to *A. bisulcatus*. Root exudates and extract from seleniferous (Pine Ridge) soil collected using these exudates were also analyzed for Se speciation. Selenium distribution and local speciation were determined using μXRF mapping and Se K-edge μXANES spectroscopy, respectively, both as described by (Quinn et al. 2011). Owing to the time-intensive nature of μXRF and μXANES studies, one biological replicate was analyzed per treatment; for each of these replicates XANES spectra were collected at three different locations on the sample. Red selenium (white line position set at 12 660 eV) was used to calibrate the spectra. Linear least-squares (LSQ) fitting of XANES spectra was performed in the 12 630–12 850 eV range using a library of standard selenocompounds. The best LSQ fit was obtained for minimum normalized sum-squares residuals: \( \text{NSS} = 100 \times \left( \frac{\sum (\mu_{\text{exp}} - \mu_{\text{fit}})^2}{\sum (\mu_{\text{exp}})^2} \right) \), where \( \mu \) represents the normalized absorbance. The error on the percentages of species present is estimated to ± 10%. All data processing and analyses were performed with a suite of custom

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Cocultivation experiment on Turface supplied with different Se concentrations

Essentially the same experimental outline was followed as described earlier for the soil cocultivation experiments, with the difference that the plants were cultured in 100% Turface growth medium, and treated once a week with different concentrations of Se (0, 10, 20, 40 or 80 μM Na₂SeO₄), and twice a week with 0.5-strength Hoagland solution. Also, five (rather than six) replicates were planted for each of the seven plant species combinations and Se concentration. At harvest, the youngest mature leaf was collected from *A. drummondii* and *S. elata* and immediately flash-frozen using liquid nitrogen for X-ray microprobe analyses as described later.

For root exudate collection, we used a modification of the method described by Cakmak et al. (1996). Specifically, plants of all four species were grown on Turface with two plants of the same species per pot \( (n = 3) \) and treated with 20 μM Na₂SeO₄ as described earlier. The plants were harvested after 6 months, gently washed, and transferred to 50 ml of distilled water. After 2 d the plants were transferred to another container with 50 ml water. After another 3 d, this second volume of water and root-released compounds (which will hereafter be referred to as exudate) was collected and analyzed for Se concentration as described in the following. Furthermore, some of the exudate fractions were frozen for Se speciation as described later. In addition, the exudate fractions were used to extract some Pine Ridge Natural Area soil. To 2 g of soil was added 6 ml of exudate, and after mixing by rotation for 1 h at room temperature, allowed to settle overnight at 4°C. The liquid fraction was then removed and used for elemental analysis and X-ray microprobe analyses as described later.
LabVIEW (National Instruments, Austin, TX, USA) programs available at the beamline.

Elemental analysis

Entire youngest mature leaves were collected from *A. bisulcatus*, *S. pinnata*, *A. drummondii* and *S. elata* for Se concentration analysis. Samples were rinsed with distilled water to remove any external Se and then dried at 45°C for 48 h. The samples were then digested in nitric acid as described by Zarcinas et al. (1987). Soil samples were dried, sieved, and extracted with ammonium bicarbonate-diethylenetriaminepentaacetic acid (AB-DTPA) as described by Soltanpour & Schwab (1977). Inductively coupled plasma atomic emission spectroscopy (ICP-AES) was used as described by Fassel (1978) to determine elemental concentrations.

Statistical analysis

The software JMP-IN (3.2.6, SAS Institute, Cary, NC, USA) was used for statistical data analysis. A Student’s *t*-test was used to compare differences between two means. ANOVA (one-way and two-way) followed by a post-hoc Tukey–Kramer test was used when comparing multiple means. In cases where the assumptions underlying these tests (normal distribution, equal variance) were not met, the data were transformed (log10, square root, or reciprocal) and reanalyzed. In one case transformation was not sufficient to meet the assumptions and therefore a non-parametric test was used (Kruskal–Wallis). Correlation analysis was used to correlate plant biomass with substrate Se concentration.

Results

When pairs of plants from the same species were grown together in one pot, the total biomass attained by Se hyperaccumulators *A. bisulcatus* and *S. pinnata* was two- to threefold larger on seleniferous (PR) soil than on nonseleniferous (CP) soil (Fig. 1a; see Table 1 for soil properties). For nonhyperaccumulator species *A. drummondii* and *S. elata*, there was no significant difference in growth on nonseleniferous and seleniferous soil (Fig. 1a). When growth on each soil was compared between the four plant species, there was a pronounced difference between hyperaccumulators and nonaccumulators with respect to their performance on seleniferous soil: the average shoot and root DW of *A. bisulcatus* and *S. pinnata* was two- to fourfold larger than those of *A. drummondii* and *S. elata* (Fig. 1b). On nonseleniferous soil there were no significant differences in growth between hyperaccumulators and nonhyperaccumulators (Fig. 1c).

When two plants from different species, one hyperaccumulator and one nonhyperaccumulator from the same genus, were grown together in one pot on seleniferous soil, the hyperaccumulators were bigger than the nonhyperaccumulators in both cases (Fig. 2a,b). *A. bisulcatus* was two- to threefold larger than *A. drummondii* (Fig. 2a); the root DW of hyperaccumulator *S. pinnata* was twofold larger than that of *S. elata*; the shoot DW was not significantly different (*P* = 0.08, Fig. 2b). Fig. 2(c–f) shows the biomass of each of the four species on seleniferous soil as influenced by which neighbor was in the same pot. The shoot and root biomass of hyperaccumulator *A. bisulcatus* was the same when grown with another *A. bisulcatus* plant as it was when it grown with nonhyperaccumulator *A. drummondii*; however, the *A. bisulcatus* biomass was twofold smaller when grown with hyperaccumulator *S. pinnata* (Fig. 2c). The shoot and root biomass of hyperaccumulator *S. pinnata* was larger when grown with another *S. pinnata* plant than when grown with nonhyperaccumulator *S. elata* or with hyperaccumulator *A. bisulcatus*. 

![Fig. 1](image.png)
Table 1 Soil properties (0–5 cm depth) at the study sites, Pine Ridge Natural Area and Cloudy Pass, Fort Collins, CO, USA

<table>
<thead>
<tr>
<th>Sample ID</th>
<th>Pine Ridge</th>
<th>Average ± SE</th>
<th>Cloudy Pass</th>
<th>Average ± SE</th>
</tr>
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<tbody>
<tr>
<td>Texture</td>
<td>Sandy Loam</td>
<td></td>
<td>Sandy Loam</td>
<td></td>
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<tr>
<td>pH</td>
<td>7.57 ± 0.03a</td>
<td>6.57 ± 0.03b</td>
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<tr>
<td>EC (mmhos cm⁻¹)</td>
<td>0.43 ± 0.03a</td>
<td>0.4 ± 0a</td>
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<td>SOM (%)</td>
<td>5.8 ± 0.06a</td>
<td>4.5 ± 0.06b</td>
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<tr>
<td>CaCO₃ (%)</td>
<td>16.25 ± 0.14a</td>
<td>2.03 ± 0.07b</td>
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<tr>
<td>NO₃-N (mg kg⁻¹)</td>
<td>2.83 ± 0.03a</td>
<td>1.23 ± 0.03b</td>
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<td>Al (mg kg⁻¹)</td>
<td>0.78 ± 0.02a</td>
<td>1.63 ± 0.09b</td>
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<td>Ba (mg kg⁻¹)</td>
<td>0.72 ± 0.02a</td>
<td>1.67 ± 0.03b</td>
<td></td>
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<tr>
<td>Ca (mg kg⁻¹)</td>
<td>350.20 ± 1.3a</td>
<td>324.30 ± 2.3b</td>
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<td>Cd (mg kg⁻¹)</td>
<td>0.41 ± 0.01a</td>
<td>0.16 ± 0.01b</td>
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<td>Cr (mg kg⁻¹)</td>
<td>0.08 ± 0.06a</td>
<td>0.11 ± 0.05a</td>
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<td>Cu (mg kg⁻¹)</td>
<td>11.7 ± 0.09a</td>
<td>4.60 ± 0.12b</td>
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<tr>
<td>Fe (mg kg⁻¹)</td>
<td>7.5 ± 0.09a</td>
<td>39.73 ± 0.12b</td>
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<td>K (mg kg⁻¹)</td>
<td>453 ± 1.74a</td>
<td>394 ± 1.53b</td>
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<tr>
<td>Mg (mg kg⁻¹)</td>
<td>61.3 ± 0.46a</td>
<td>149 ± 1.97b</td>
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<tr>
<td>Mn (mg kg⁻¹)</td>
<td>11.67 ± 0.09a</td>
<td>4.57 ± 0.09b</td>
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<tr>
<td>Mo (mg kg⁻¹)</td>
<td>0.04 ± 0.01a</td>
<td>0.01 ± 0b</td>
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<td>Na (mg kg⁻¹)</td>
<td>24.40 ± 0.25a</td>
<td>13.97 ± 0.12b</td>
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<td>Ni (mg kg⁻¹)</td>
<td>1.15 ± 0.02a</td>
<td>0.42 ± 0.02b</td>
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<td>P (mg kg⁻¹)</td>
<td>2.1 ± 0.06a</td>
<td>8.1 ± 0.06b</td>
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<td>Pb (mg kg⁻¹)</td>
<td>1.6 ± 0.06a</td>
<td>1.23 ± 0.09b</td>
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<td>S (mg kg⁻¹)</td>
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<td>15.63 ± 0.12b</td>
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<td>Se (mg kg⁻¹)</td>
<td>1.7 ± 0.06a</td>
<td>0.11 ± 0b</td>
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<td>V (mg kg⁻¹)</td>
<td>1.2 ± 0.06a</td>
<td>0.15 ± 0b</td>
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<tr>
<td>Zn (mg kg⁻¹)</td>
<td>1.63 ± 0.12a</td>
<td>2.33 ± 0.03b</td>
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Shown are means ± standard error (n = 3). a and b denote significant differences.

(Fig. 2d). The shoot and root biomass of nonhyperaccumulator A. drummondii was twofold lower when grown next to another A. drummondii than when grown next to hyperaccumulator A. bisulcatus (Fig. 2c). The shoot biomass of nonhyperaccumulator S. elata was significantly smaller when grown next to A. bisulcatus (Fig. 2e). The shoot biomass of nonhyperaccumulator S. elata was bigger than when grown next to S. pinnata; the root biomass was not significantly different (Fig. 2f).

When a hyperaccumulator and a nonhyperaccumulator from the same genus were cocultivated on nonseleniferous soil, the nonhyperaccumulator A. drummondii was twofold bigger than the hyperaccumulator A. bisulcatus (Fig. 3a), while the two Stanleya species did not show significantly different growth (Fig. 3b, P = 0.136 for shoot and 0.076 for root). Interestingly, the average shoot and root DWs of hyperaccumulator A. bisulcatus were twofold higher when grown with hyperaccumulator S. pinnata than when grown with another A. bisulcatus or with nonhyperaccumulator A. drummondii (Fig. 3c). Similarly, there was a pronounced increase in the average shoot and root DWs of hyperaccumulator S. pinnata when it was grown with hyperaccumulator A. bisulcatus compared with when it was grown with another S. pinnata plant or with nonhyperaccumulator S. elata (Fig. 3d). Nonhyperaccumulator A. drummondii was bigger when grown next to A. bisulcatus than when grown next to another A. drummondii (Fig. 3e). Nonaccumulator S. elata shoots were significantly bigger when growing next to hyperaccumulator S. pinnata than when growing next to S. elata; there was no difference in root biomass (Fig. 3f).

Two-way ANOVA was used to test whether the relationship between a pair of plants was different in high-Se vs low-Se soil. For A. bisulcatus a significant interaction was found for both shoot and root DW (P < 0.0001): A. bisulcatus plants that had S. pinnata as a neighbor had less biomass when grown on high-Se soil, but more biomass when grown on low-Se soil, as compared with plants that had other neighbors (Figs 2c, 3c). There was also a significant interaction (P < 0.0001) for both shoot and root DW for S. pinnata plants that had S. pinnata as a neighbor had more biomass when grown on high-Se soil, but less biomass on low-Se soil compared with plants with other neighbors (Figs 2d, 3d). The effects of neighboring species on the growth of the two nonhyperaccumulator species were less soil-dependent. Only the (positive) effect of S. pinnata on root DW of S. elata was soil-dependent, namely only observed on low-Se soil.

Fig. 4 shows the Se concentration in the shoots and roots of each of the four species on seleniferous soil, as influenced by which neighbor was in the same pot. The most pronounced neighbor effect was found for hyperaccumulator S. pinnata, whose shoot and root Se concentration was eight to 10-fold higher when grown in a pot with nonhyperaccumulator S. pinnata than when grown with another S. pinnata or with hyperaccumulator A. bisulcatus (Fig. 4b). Furthermore, roots of A. bisulcatus contained a lower Se concentration when growing next to S. pinnata compared with other species (Fig. 4a), and roots of A. drummondii contained a lower Se concentration when growing next to A. bisulcatus than when growing next to A. drummondii (Fig. 4c). Nonsignificant trends worth noting are that the Se concentration in A. bisulcatus was c. 50% higher when growing next to another A. bisulcatus than when growing next to A. drummondii or S. pinnata (Fig. 4a), and the Se concentration in S. pinnata was c. 50% higher when growing next to another S. pinnata than when growing next to A. bisulcatus (Fig. 4b). As a result of the combined neighbor effects on plant growth and Se concentration, the total shoot Se accumulation per plant, as calculated from the product of shoot Se concentration (Fig. 4) and shoot biomass (Fig. 2), was threefold higher when the hyperaccumulators were grown next to a hyperaccumulator from the same species as compared with a hyperaccumulator of the other species (P = 0.0005 for A. bisulcatus and 0.036 for S. pinnata). A similar calculation can be done for S. elata, whose shoot Se concentration was 50% higher when grown next to S. pinnata than S. elata (Fig. 4d, P = 0.208); the total shoot Se accumulation per S. elata plant was threefold higher next to S. pinnata (P = 0.088).

The Se concentrations in shoots and roots of the four species when grown on nonseleniferous soil are shown in Fig. 5. As expected, the Se concentrations were substantially lower than in the plants grown on seleniferous soil, but still measurable. The shoot Se concentration in hyperaccumulator A. bisulcatus was two- to threefold higher when growing in the same pot with nonhyperaccumulator A. drummondii or with hyperaccumulator S. pinnata than when growing with another A. bisulcatus plant; there were no differences in root Se concentration (Fig. 5a). The root and shoot Se concentrations in hyperaccumulator S. pinnata
were five to 10-fold higher when grown in same pot with non-hyperaccumulator *S. elata* than when grown with another *S. pinnata* or with hyperaccumulator *A. bisulcatus* (Fig. 5b), which is similar to the results found on seleniferous soil (Fig. 4b). Root Se concentration of nonhyperaccumulator *A. drummondii* was twofold lower when growing in a pot with another *A. drummondii* than when growing with hyperaccumulator *A. bisulcatus* (Fig. 5c). There were no significant differences in shoot Se concentration \((P = 0.36, \text{Fig. } 5c)\), or in total Se accumulated per *A. drummondii* shoot, which was 2-2 fold higher when *A. bisulcatus* was the neighbor \((P = 0.18)\). *S. elata* root Se concentrations were about fivefold lower when growing next to *S. elata* than when grown next to *S. pinnata* (Fig. 5d, \(P < 0.05\)). The shoot Se concentrations did not differ significantly \((P = 0.31, \text{Fig. } 5d)\), but the total shoot Se accumulation per *S. elata* plant (calculated from the product of Se concentration and biomass) was threefold higher next to *S. pinnata* than when next to *S. elata* \((P = 0.066)\).

The relationship between pairs of plants was in some cases different in high-Se and low-Se soils, as determined by two-way ANOVA. *A. bisulcatus* plants that had *S. pinnata* as a neighbor had lower root Se concentrations than *A. bisulcatus* plant that had other neighbors, but only on high-Se soil (Figs 4c, 5c; \(P = 0.011\)). There was also a significant interaction for both shoot and root Se concentrations for *S. pinnata*: having *S. elata* as a neighbor had a significantly bigger positive effect on Se concentration in *S. pinnata* when grown on high-Se soil than when grown on low-Se soil (Figs 4d, 5d; \(P < 0.001\)). Furthermore, root Se concentration in *A. drummondii* was affected positively by its neighbor *A. bisulcatus* on low-Se soil, but negatively on high-Se soil \((P < 0.01)\).

To be able to separate the effect of Se on plant–plant interactions from that of other factors (other soil properties, microbial composition), a second cocultivation experiment was carried out using Turface growth medium supplied with different concentrations of Na2SeO4. When pairs of plants from the same species were grown together in one pot, there was an opposite growth response in Se hyperaccumulators and nonaccumulators. Total plant biomass showed a positive correlation \((P < 0.05)\) with...
increasing external Se concentration for hyperaccumulators *A. bisulcatus* and *S. pinnata*, being 4.5-fold and twofold bigger, respectively, when treated with 80 μM Na₂SeO₄ than in the absence of Se (Fig 6a,b). By contrast, the biomasses of nonhyperaccumulators *A. drummondii* and *S. elata* decreased six- and 15-fold, respectively, with increasing Se concentration (*P* < 0.05, Fig 6c,d). Thus, the hyperaccumulators were not only Se-tolerant, but even benefited from increasing Se supply, while the nonaccumulators were Se-sensitive, showing 50% growth inhibition at external Se concentrations between 5 and 15 μM.

When two plants from different species, one hyperaccumulator and one nonhyperaccumulator from the same genus, were grown together in one pot, similar growth responses to Se were observed. Hyperaccumulator *A. bisulcatus* had a threefold increase in size with increasing Se treatment, while the cocultivated nonhyperaccumulator *A. drummondii* decreased > 10-fold in size (Fig. 6e). *Stanleya pinnata* had a twofold increase in size with increasing Se supply while nonhyperaccumulator *S. elata* decreased over 60-fold in size (Fig. 6f). As a result of these differential growth responses to Se, the nonaccumulators were bigger than the hyperaccumulators in the absence of Se, while the hyperaccumulators outgrew the nonaccumulators above external Se concentrations of 3 and 8 μM Na₂SeO₄, respectively, for the *Astragalus* and *Stanleya* pairs (Fig. 6e,f). When the two hyperaccumulator species were grown together in one pot, their growth responses were also similar to those observed when grown individually: the biomasses of *A. bisulcatus* and *S. pinnata* increased 2.5-fold and fourfold, respectively, with increasing Se supply (Fig. 6g).

![Fig. 3](https://example.com/f3.png) Shoot and root biomass (g DW) of hyperaccumulator plants *Astragalus bisulcatus* (*A. bis*) and *Stanleya pinnata* (*S. pin*) and nonaccumulators *Astragalus drummondii* (*A. dru*) and *Stanleya elata* (*S. ela*) grown in pots on nonseleniferous soil from Cloudy Pass. Two plants from the same or different species were grown per pot. (a,b) Biomass of each of two neighbors that were cocultivated in one pot. (c–f) Growth of each of the four species as influenced by which neighbor was in the same pot. Values shown are means ± SE (*n* = 6); different lower-case letters above the bars indicate significantly different means (ANOVA, *α* = 0.05).
nonhyperaccumulator *S. elata* than when growing with another *S. pinnata* (Fig. 7b). Nonhyperaccumulators *A. drummondii* and *S. elata* showed increasing tissue Se concentration with increasing Se supply, which was similar in plants grown with a hyperaccumulator or a nonaccumulator neighbor (Fig. 7c,d).

To obtain a better understanding of the mechanism responsible for the observed effects of neighboring plants on plant Se accumulation, root exudate was collected from each of the four species after being grown on Turface and treated with 20 μM Na₂SeO₄. The shoot and root Se concentrations in

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**Fig. 4** Selenium concentration (mg kg⁻¹ DW) in shoot and root of hyperaccumulators *Astragalus bisulcatus* (*A. bis*) (a) and *Stanleya pinnata* (*S. pin*) (b) and nonaccumulators *Astragalus drummondii* (*A. dru*) (c) and *Stanleya elata* (*S. ela*) (d) after being grown in pots on seleniferous soil from Pine Ridge Natural Area with either another plant from the same species or one from a different species as neighbor. Values shown are means ± SE (*n* = 6); different lower-case letters above the bars indicate significantly different means (ANOVA, *α* = 0.05).

**Fig. 5** Selenium concentration (mg kg⁻¹ DW) in shoot and root of hyperaccumulators *Astragalus bisulcatus* (*A. bis*) (a) and *Stanleya pinnata* (*S. pin*) (b) and nonaccumulators *Astragalus drummondii* (*A. dru*) (c) and *Stanleya elata* (*S. ela*) (d) after being grown in pots on non-seleniferous soil from Cloudy Pass with either another plant from the same species or one from a different species as neighbor. Values shown are means ± SE (*n* = 6); different lower-case letters above the bars indicate significantly different means (ANOVA, *α* = 0.05).
hyperaccumulators *A. bisulcatus* and *S. pinnata* were, on average, two- to threefold higher than those in nonhyperaccumulators *A. drummondii* and *S. elata* grown on Turface (gravel) growth medium and treated with different concentrations of Na$_2$SeO$_4$. (a–d) Two plants from the same species grown in one pot; (e–g) Two plants from different species grown in one pot: (e) *A. bisulcatus* (closed circles) and *A. drummondii* (open circles); (f) *S. pinnata* (closed circles) and *S. elata* (open circles); (g) *A. bisulcatus* (closed circles) and *S. pinnata* (open circles). Values shown are means ± SE ($n = 5$); different lower-case letters above bars indicate significantly different means (ANOVA, $\alpha = 0.05$).
In addition to affecting the total Se concentration in neighboring plants, it is also feasible that plants can affect their neighbor’s Se speciation (i.e. the chemical composition of the selenocompounds). To investigate the Se speciation in nonaccumulator species *A. drummondii* and *S. elata* as affected by their neighbor in the same pot, Se K-edge XANES spectra were collected in leaves of plants grown on Turface and treated with $20 \mu M$ $Na_2SeO_4$ (Table 2). The Se in both nonaccumulators consisted primarily (89–95%) of an organic C-Se-C compound, indistinguishable from the standards selenomethionine and methyl-selenocysteine; the remainder was selenate ($SeO_4^{2-}$) (Table 2). The relative abundance of C-Se-C and selenate were similar in *A. drummondii* leaves collected from plants growing next to *A. drummondii* or growing next to *A. bisulcatus*. However, speciation in *S. elata* leaves was different when its neighbor was *S. elata* than when its neighbor was *S. pinnata*. *S. elata* that was grown next to *S. pinnata* showed a 3.5-fold lower ($P < 0.01$) selenate fraction and a concomitant...
A. drummondii grown next to A. drummondii
Spot 1 7.8 95.8 5.8 ND
Spot 2 10.9 96.0 5.9 ND
Spot 3 8.9 90.0 8.9 ND
Average ± SE 93.9 ± 1.9 6.8 ± 1.0 ND

A. drummondii grown next to Astragalus bisulcatus
Spot 1 5.1 94.9 6.2 ND
Spot 2 5.4 93.9 6.9 0.4
Spot 3 4.5 90.8 9.5 ND
Average ± SE 93.2 ± 1.0 7.5 ± 1.0 ND

Stanleya elata grown next to S. elata
Spot 1 5.1 88.7 9.8 ND
Spot 2 4.0 89.7 11.0 ND
Spot 3 4.7 88.3 11.9 ND
Average ± SE 88.9 ± 0.3 10.9 ± 0.6 ND

S. elata grown next to Stanleya pinnata
Spot 1 3.5 94.0 3.7 ND
Spot 2 4.0 95.4 3.5 ND
Spot 3 3.4 96.4 2.4 ND
Average ± SE 95.2 ± 0.6 3.2 ± 0.6 ND

A. bisulcatus root exudate
Spot 1 25.6 70.7 ND 33.5
Spot 2 6.4 90.6 ND 11.9
Spot 3 11.6 96.7 ND 4.5
Average ± SE 86 ± 8.0 ND 16.6 ± 8.8

Table 2 Selenium speciation in plant leaf material and root exudates determined from X-ray absorption near edge structure (XANES) linear least-squares (LSQ) fitting

Plants were grown in pairs on Turface (gravel) and supplied with selenate. One plant per treatment was analyzed and three spectra were obtained from each plant. Values shown for each form of Se represent the % of total Se. The best LSQ fit was obtained for minimum normalized sum-squares residuals (NSS), NSS = 100 × (Σ(μexp - μfit)²/Σ(μexp)²) where μ represents the normalized absorbance. ND, not detectable; C-Se-C, MeSeCys, SeMet, SeCysteine (indistinguishable). Forms of Se that were not detected in any of the samples and therefore not tabulated: red and gray Se⁰, Se-cystine, Se-cysteine, Se(GSH)₂.

increase in C-Se-C abundance (P < 0.001) compared with S. elata grown next to another S. elata (Fig. 9, Table 2). The Se speciation in the root exudates and soil extracts obtained using root exudates was also analyzed by XANES. Only the A. bisulcatus exudate provided useful Se spectra; the main selenocompound (83%) in the exudate was organic Se of a C-Se-C type, and the remainder was selenite (Table 2).

Discussion
The finding that Se hyperaccumulators perform better on seleniferous soil may indicate that they benefit physiologically from Se. Indeed, the Turface experiment showed that the two hyperaccumulators grew several-fold better with increasing Se concentration, suggesting that the Se in the seleniferous soil was responsible for the better growth of the hyperaccumulators. The beneficial effect of Se on hyperaccumulator growth was previously described by Shrift (1969). A possible mechanism may be enhanced antioxidant activity, as was found for nonhyperaccumulator species (Cartes et al., 2005; Hartikainen, 2005).

In the Turface experiment the growth of the nonaccumulators was worse when Se supply increased, reaching 50% inhibition at 10–20 μM sodium selenate (0.8–1.6 ppm Se), corresponding with a tissue Se concentration of c. 200–250 mg kg⁻¹ DW. This is similar to what was found earlier for Arabidopsis thaliana (El Mehdawi et al., 2011a). However, the form of Se in A. drummondii and S. elata was mainly organic C-Se-C (89–95%), while other nonaccumulator species, including A. thaliana, accumulate mainly selenite with a minor fraction of C-Se-C (de Souza et al., 1998; Van Hoewyk et al., 2005; Freeman et al., 2006a). Based on XANES data alone the C-Se-C compound in A. drummondii and S. elata could be MeSeCys, SeMet or SeCysth; these cannot be distinguished. MeSeCys was found earlier to be the predominant form of Se in hyperaccumulators S. pinnata and A. bisulcatus, which explains their Se tolerance, since MeSeCys does not enter proteins. The intermediate Se accumulator Stanleya albescens, on the other hand, accumulated mainly SeCysth and was fairly Se-sensitive (Freeman et al., 2010). The Se sensitivity in A. drummondii and S. elata could be a result of the accumulation of the more toxic forms SeMet or SeCysth, or of the fact that the remainder of their Se was selenite (4–11%). This form of Se is toxic when accumulated, as a result of pro-oxidant activity (Grant et al., 2011).

The opposite growth responses to Se may affect competition between hyperaccumulators and nonaccumulators: the two likely have different competitive strength depending on soil Se concentrations. When cocultivated in Turface at different Se
In seleniferous soil the Se concentrations were much lower than the Se release from hyperaccumulators. This was found on seleniferous soil, on nonseleniferous soil, as well as in Turface. The mechanism for this positive effect is not readily apparent. \textit{S. elata} roots were shown to release some Se, but these concentrations were much lower than the Se release from hyperaccumulators. It was interesting, however, that the \textit{S. elata} exudate extracted twofold more Se from seleniferous soil than the \textit{S. pinnata} exudate. Thus, \textit{S. elata} exudate may somehow enhance Se bioavailability for \textit{S. pinnata}. The mechanism is not clear but could, for instance, involve a Se chelator.

The positive effect (threefold) of \textit{S. pinnata} on total shoot Se accumulation in \textit{S. elata} on both soils \((P < 0.1)\) might be caused by root Se release, as observed in \textit{S. pinnata} exudate. On Turface there was no effect of \textit{S. pinnata} on \textit{S. elata} growth or Se accumulation. However, \textit{S. pinnata} did seem to affect Se speciation in \textit{S. elata}. \textit{S. elata} contained relatively more organic Se when its neighbor was \textit{S. pinnata} than when it was another \textit{S. elata}. This may be the result of root release of organic Se by \textit{S. pinnata}. In support of this hypothesis, \textit{S. pinnata} roots were shown here to exude significant concentrations of Se, and the form of Se in roots of \textit{S. pinnata} was shown recently to be C-Se-C (Lindblom et al., 2011). While the exudate of \textit{S. pinnata} did not have a strong enough Se signal to obtain reliable speciation information from XANES, \textit{A. bisulcatus} exudate was shown by XANES to contain predominantly C-Se-C. Thus, hyperaccumulators may exude organic Se and, since the main form of bioavailable Se in soil is thought to be inorganic selenate, the root release of organic Se may affect local Se speciation, and with that, Se bioavailability and Se uptake and speciation by neighboring plants. Enhanced bioavailability of Se around hyperaccumulators was also suggested by the earlier finding that, while the soil around hyperaccumulators was seven- to 13-fold enriched with Se, the neighboring plants were enriched up to 20-fold (El Mehdawi et al., 2011b). The finding that hyperaccumulators release Se from their roots supports the hypothesis that hyperaccumulators can phytoenrich their surrounding soil with Se, and that root release of Se is one of the mechanisms for phytoenrichment. Litter deposition and decomposition likely is another mechanism, as indicated by an earlier study (Quinn et al., 2010).

Hyperaccumulator \textit{A. bisulcatus} had a positive effect on growth of nonaccumulator \textit{A. drummondii} on both soils. The neighbor effects between the \textit{Astragalus} species with respect to Se accumulation varied. On nonseleniferous soil where Se concentrations were very low, \textit{A. bisulcatus} plants growing next to \textit{A. drummondii} contained fivefold higher Se concentrations than \textit{A. bisulcatus} plants growing next to \textit{A. bisulcatus}; \textit{A. drummondii} also contained elevated Se concentrations when growing next to \textit{A. bisulcatus} compared with another \textit{A. drummondii}. No such effects were seen on seleniferous soil, but on Turface \textit{A. drummondii} also appeared to stimulate Se accumulation in \textit{A. bisulcatus} at all Se concentrations. The positive effect of \textit{A. drummondii} on Se uptake by \textit{A. bisulcatus} when external Se concentrations were low may be a result of the release of Se chelators by \textit{A. drummondii} roots, whose presence is suggested from the observation that \textit{A. drummondii} exudate did not contain much Se but released more Se from soil than \textit{A. bisulcatus} exudate.

The two hyperaccumulators \textit{A. bisulcatus} and \textit{S. pinnata} affected each other's growth and Se accumulation negatively on seleniferous soil, but positively on nonseleniferous soil. This opposite effect may be a result of the 17-fold different Se concentrations, as well other soil properties. Different elements may limit hyperaccumulator growth on the two soils, and the limiting factors may be different for the two hyperaccumulator species. Iron and phosphorous concentrations were four- to fivefold lower in Pine Ridge soil, while nitrate concentrations were twofold higher than in Cloudy Pass soil (Table 1); these elements are often limiting for plant growth, and therefore may have affected plant growth and competition. Furthermore, the Pine Ridge soil was slightly basic while the Cloudy Pass soil was slightly acidic, and soil organic matter was somewhat higher at Pine Ridge, which may have further affected nutrient concentration and bioavailability. In addition to affecting plants directly, these soil properties may affect plant–plant and plant–microbe interactions, for instance by affecting the bioavailability of exuded selenocompounds or Se chelators. Based on the limited information available, we can only speculate about which factors may have caused the opposite effects of the hyperaccumulators on each other. If on Pine Ridge soil Se would be limiting hyperaccumulator growth, and the two species use different mechanisms (e.g. different chelators) to make the soil Se bioavailable, then two plants of the same species may facilitate each other’s growth by working together in making the Se bioavailable (as indicated by the higher plant Se concentrations), while two plants of different species may cause the limited Se less available for the other species. On the nonseleniferous soil, Se may not play a role in plant competition and other elements such as N, Fe, P or K may be limiting plant growth. If the two different hyperaccumulator species are limited by different elements, two plants of the same

species may impede each other’s growth by competing for the same limiting nutrient, while two plants of different species may be more complementary in their nutrient requirements, and therefore inhibit each other’s growth to a lesser degree.

These studies provide better understanding of the role of Se in plant–plant interactions, particularly between hyperaccumulators and nonaccumulators. Hyperaccumulator species perform better on seleniferous than on nonseleniferous soil, and in general grow better with increasing Se supply. Growth of nonaccumulator species gets worse with increasing Se concentrations. It appears that both hyperaccumulators and nonaccumulators can affect the growth and Se accumulation of neighboring plants. Roots of hyperaccumulators can exude significant concentrations of Se, mainly in organic form, which may lead to higher fractions of organic Se in nonaccumulator neighbors. Nonaccumulators, on the other hand, may be able to enhance soil Se bioavailability, and, with that, Se concentrations in their neighbors. These results are of significance since they offer an insight into how Se affects competition and facilitation between plants, and why hyperaccumulators are found almost exclusively on seleniferous soils.

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